REVIEW



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Venomous snake bites: clinical diagnosis and treatment

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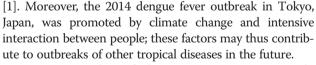
Abstract

Snake bites are life-threatening injuries that can require intensive care. The diagnosis and treatment of venomous snake bites is sometimes difficult for clinicians because sufficient information has not been provided in clinical practice. Here we review the literature to present the proper management of bites by mamushi, habu, and yamakagashi snakes, which widely inhabit Japan and other Asian countries. No definite diagnostic markers or kits are available for clinical practice; therefore, definitive diagnosis of snake-venom poisoning requires positive identification of the snake and observation of the clinical manifestations of envenomation. Mamushi (Gloydius blomhoffii) bites cause swelling and pain that spreads gradually from the bite site. The platelet count gradually decreases due to the platelet aggregation activity of the venom and can decrease to <100,000/mm³. If the venom gets directly injected into the blood vessel, the platelet count rapidly decreases to <10,000/mm³ within 1 h after the bite. Habu (Protobothrops flavoviridis) bites result in swelling within 30 min. Severe cases manifest not only local signs but also general symptoms such as vomiting, cyanosis, loss of consciousness, and hypotension. Yamakagashi (Rhabdophis tigrinus) bites induce life-threatening hemorrhagic symptoms and severe disseminated intravascular coagulation with a fibrinolytic phenotype, resulting in hypofibrinogenemia and increased levels of fibrinogen degradation products. Previously recommended first-aid measures such as tourniquets, incision, and suction are strongly discouraged. Once airway, breathing, and circulation have been established, a rapid, detailed history should be obtained. If a snake bite is suspected, hospital admission should be considered for further follow-up. All venomous snake bites can be effectively treated with antivenom. Side effects of antivenom should be prevented by sufficient preparation. Approved antivenoms for mamushi and habu are available. Yamakagashi antivenom is used as an off-label drug in Japan, requiring clinicians to join a clinical research group for its use in clinical practice.

Keywords: Mamushi, Habu, Yamakagashi, Antivenom

Introduction

Throughout the world, snake bites remain life-threatening injuries [1-4], sometimes requiring intensive care [5]. Similar to malaria, dengue hemorrhagic fever, tuberculosis, and parasitic diseases, the risk of snake bite is always present [1]. In 2009, the World Health Organization (WHO) added snake bites to the list of neglected tropical diseases, which includes dengue hemorrhagic fever, cholera, and Japanese encephalitis. The mortality associated with snake bites is much greater than that of other neglected tropical diseases



Venomous snakes of the same genus as mamushi (*Gloydius*), habu (*Protobothrops*), and yamakagashi (*Rhabdophis*) inhabit Japan and other Asian countries [6-8]. The incidence of bites by these venomous snakes is reported as approximately 1,000 cases with 10 deaths annually for mamushi (*Gloydius blomhoffii*) [9], 100 cases annually for habu (*Protobothrops flavoviridis*) [10], and 34 cases with 4 deaths over the past 40 years for yamakagashi (*Rhabdophis tigrinus*) [6].



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The diagnosis and treatment of venomous snake bites is sometimes difficult for clinicians because sufficient information, including the administration of antivenom therapy, has not been provided in clinical practice [6,11]. Here we clarify the proper management of bites by mamushi, habu, and yamakagashi, including snake characteristics, venom activity and symptoms, clinical diagnosis, and treatment.

Review

Snake characteristics Mamushi (G. blomhoffii)

Mamushi is a pit viper that is seen in a wide variety of colors (Figure 1). As mamushi is a small snake (about 60 cm), its attack range is only about 30 cm [11]. The fangs are about 5 mm long, with very thin tips (Figure 2a). This snake lives near rivers, ponds, and paddy fields and is active in the daytime in spring and autumn and at night in the summer. In Japan, *G. blomhoffii* is seen from Kyushu to Hokkaido, and the distinct species *Gloydius tsushimaensis* (Tsushima Mamushi) is found on Tsushima island, Nagasaki.

Habu (P. flavoviridis)

Five types of pit vipers inhabit Okinawa and Amami. Habu, one of these pit vipers, varies in color by region (Figure 3). Even though this nocturnal snake is not active in the daytime, many people are bitten when disturbing snakes while farming. At night, this snake comes out in search of food near houses, sometimes entering them. Accidents often occur during handling. Habu snakes often climb trees. Habu is the most dangerous of these three snakes because it is large, reaching up to 2 m in length, and is the most aggressive. Habu fangs are tubular and 1.5–2 cm in length (Figure 2b). Dry bites can occur because the venom-releasing pore of the habu snake is located approximately 0.1 cm from the tip of the venom fang [12].

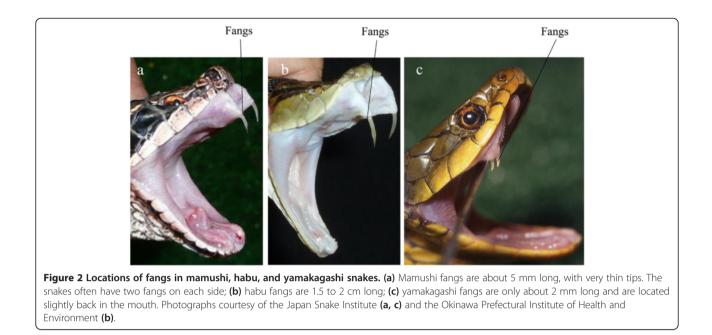
Yamakagashi (R. tigrinus)

Yamakagashi is a rear-fanged venomous snake that lives near rivers, ponds, and paddy fields, the same habitat as mamushi. Snakes of the same genus, such as *Rhabdophis lateralis* and *Rhabdophis subminiatus*, are distributed throughout Russia and Asia [13,14]. Yamakagashi grows to about 1 m in the plains and 1.5 m in the hills and mountains. The color varies by region (Figure 4). The larger snakes have short, 2-mm long fangs located slightly back from the front of the mouth. Like viper fangs, the fangs of yamakagashi are not tubular, and the venom gland duct opens at the base of the fang (Figure 2c). Because yamakagashi fangs are not grooved, envenomation does not occur in most bites; therefore, this snake has long been considered non-venomous [13,15].

Venom activity and clinical symptoms Mamushi (G. blomhoffii)

Several enzymes, including a protease, phospholipase A2 (PIA2), and bradykinin-releasing-enzyme are contained in the mamushi venom [16]. The effects of these enzymes are described in Table 1. Local pain and swelling are the main symptoms at the bite site; subcutaneous bleeding and blisters are sometimes observed. The swelling and pain spread gradually from the bite site





(Table 2). Most patients are bitten on the hand or foot, but the spread of swelling to the trunk is often observed [17].

With severe swelling, hypotension can occur. In these cases, increased levels of creatine phosphokinase (CPK) and blood myoglobin due to rhabdomyolysis are remarkable and can cause acute renal failure [11,17]. In addition to hypotension, renal hemorrhage and direct action of the

venom on the kidney can cause acute renal failure. In severe cases, the plasma potassium level can increase due to muscle tissue damage and metabolic acidosis, causing cardiac arrest shortly after the bite [18,19]. A rise in the level of the CPK isozyme cardiac muscle conformer (MB) and necrosis of the myocardium have been reported, which may be due to the direct action of venom on the cardiac muscle [20].





As the venom is absorbed from the bite site, the platelet count gradually decreases due to the platelet aggregation activity of the venom, sometimes decreasing to <100,000/ mm³ [21]. Cases in which the platelet count rapidly

| Enzyme | Mamushi | Habu | Yamakagashi |
|-----------------------------------|---------|------|-------------|
| Non-hemorrhagic metalloproteinase | + | + | + |
| HR1 | + | + | |
| HR2 | + | + | |
| Phospholipase A2 | + | + | |
| TAME esterase | + | + | |
| L-amino acid oxidase | + | + | |
| Hyaluronidase | + | + | |
| Phosphodiesterase | + | + | |
| Phosphomonoesterase | + | + | |
| ATPase | + | + | |
| 5'-nucleotidase | + | + | |
| Endonuclease | + | + | |
| NAD-nucleosidase | + | ± | |
| Alginine ester hydrolase | + | + | |
| Thrombin-like enzyme | + | + | ± |
| Endopeptidase | + | + | |
| Bradykinin-releasing enzyme | + | + | |

Table 1 Enzymes in the snake venoms

+ indicates the presence of the enzyme, – indicates the absence of the enzyme, blank space indicates unknown whether the venom contains the given enzyme, *HR 1* hemorrhagic factor-1, *HR-2* hemorrhagic factor-2, *TAME* p-toluenesulfonyl-L-arginine methyl ester, *ATPase* adenosine triphosphatase, *NAD* nicotinamide adenine dinucleotide.

decreases to <10,000/mm³ within 1 h after the bite are often seen [22]. The venom is thought to be injected directly into the blood vessel during the bite, as the tips of the mamushi fangs are very thin. The platelet aggregation and hemorrhagic activities are very strong, causing ecchymosis and gastrointestinal bleeding. However, even in severe cases, little change in prothrombin time (PT), activated partial thromboplastin time (APTT), or fibrinogen levels is observed [11]. The vasodilatation activity of the venom is strong, sometimes causing hypotension [23].

The venom contains small amounts of neurotoxin, which causes diplopia, blurred vision, and a divergent squint due to action on the nervus oculomotorius, but respiratory muscle paralysis is not seen. These ocular symptoms remit within several days to about 2 weeks [24].

Habu (P. flavoviridis)

The toxicity of habu venom is about half that of mamushi venom, but the amount of habu venom is approximately 10 times that of mamushi venom. Since habu venom contains many enzymes similar to those found in mamushi venom (except the neurotoxin), a similar range of symptoms are observed in patients with habu bites (Table 1). Habu venom causes extreme local swelling, necrosis, and bleeding at the bite site (Table 2). Most habu bites start to swell within 30 min after the bite [7]. In addition, severe cases manifest not only with local signs but also with general symptoms such as vomiting, cyanosis, loss of consciousness, and hypotension. Habu bites frequently cause compartment syndrome (CS) because of the large volume of venom injected, regardless of its lower

| Table 2 Typica | I symptoms and | laboratory c | lata to | be evaluated |
|----------------|----------------|--------------|---------|--------------|
|----------------|----------------|--------------|---------|--------------|

| | Mamushi | Habu | Yamakagashi |
|--|---|---|--|
| Typical symptoms | Local pain, swelling, severely decreased platelet count ^a , diplopia, blurred vision, nausea ^b , vomiting ^b , stomachache ^b , diarrhea ^b , cyanosis ^b | Local swelling, necrosis, bleeding at the bite site, vomiting ^b , cyanosis ^b , loss of consciousness ^b , hypotension ^b , compartment syndrome | Nasal bleeding, gum bleeding, bleeding from the bite site, headache ^b |
| Laboratory data to be evaluated routinely | CBC, CK, BUN, Cre, Na, K, Cl, Fibrinogen, FDP, d-din | ner, PT, APTT | |
| Typical laboratory findings | CK↑ | | FDP >100 µg/mL |
| | Plt <10,000/µL ^a | | Fibrinogen <100 mg/dL |
| Laboratory data to be evaluated additionally | Myoglobin, CK-MB | Myoglobin | AT-III, TAT, PIC |

CBC complete blood count, *Plt* platelet count, *CK* creatine kinase, *BUN* blood urea nitrogen, *Cre* creatinine, *FDP* fibrinogen degradation products, *PT* prothrombin time, *APTT* activated partial thromboplastin time *AT-III* antithrombin III, *TAT* thrombin-antithrombin III complex, *PIC* plasmin-a2-plasmin inhibitor complex. ^aVenom is injected into the blood vessel directly.

^bIn severe cases.

toxicity compared to mamushi venom. In addition, following the bite, patients tend to bind the wound excessively tightly due to fear of the venom spreading to the whole body, thereby exacerbating CS. Thus, many cases of CS are reported following habu bites.

Yamakagashi (R. tigrinus)

Yamakagashi venom (metalloproteinase) has strong blood coagulation activity, with a prothrombin-activating effect and a weak thrombin-like effect [25]. Once yamakagashi venom enters the blood, it activates prothrombin continuously, causing excessive coagulation. Disseminated fibrin formation ensues, and fibrinolysis is activated, resulting in hypofibrinogenemia and increased levels of fibrinogen degradation products (FDP) [5]. This venom induces life-threatening hemorrhagic symptoms and severe disseminated intravascular coagulation (DIC) with a fibrinolytic phenotype that is typically observed in patients with acute, severe blunt trauma [26], acute leukemia (particularly in acute promyelocytic leukemia) [27], and massive obstetric hemorrhage [28]. DIC progresses to acute renal failure due to the obstruction of glomeruli by thrombi. Because the fangs of this snake are very short, the venom is injected subcutaneously or intradermally. However, pain, swelling, and inflammation are minimal at the bite site because the venom does not act on the tissues directly. The typical symptom is hemorrhage, including nasal bleeding, gum bleeding, and bleeding from the bite site (Table 2). In severe cases, headache is also a characteristic symptom [5].

Diagnosis

There are no definite diagnostic markers or kits available in clinical practice; therefore, definitive diagnosis of snakevenom poisoning requires positive identification of the snake and observation of the clinical manifestations of envenomation [3]. On initial assessment, CBC, BUN, Cre, Na, K, Cl, CK, and coagulation markers (fibrinogen, FDP, d-dimer, PT, and APTT) should be examined (Table 2).

Mamushi (G. blomhoffii)

Because mamushi hide in the grass and fallen leaves, identification is difficult, even in the daytime. Patients usually feel only a pain similar to that of a splinter because the fangs are about 5 mm long and very thin. Thus, patients and physicians often mistake this bite for an insect bite or sting, especially when bitten at night [11]. The mamushi bite usually leaves two very small wounds that are 1 cm apart [11]. These snakes often have two fangs on each side; therefore, three or four fang marks are often observed. As small bite marks may be difficult to observe, diagnosis by bite wounds alone is difficult [11]. In many cases, blood test data do not change for several hours after the bite. If symptoms such as swelling are seen, it is necessary to perform frequent blood tests. With increased swelling, CK and blood myoglobin levels rise, followed by a rise in BUN and creatinine levels. A remarkable rise in myoglobin level is an indicator for the diagnosis of mamushi bite and suggests the risk of acute renal failure.

In cases where venom was injected into the blood vessel directly, platelet counts rapidly decrease to <10,000/mm³ but fibrinogen levels do not decrease [22]. Such cases are difficult to diagnose because the local symptoms are mild. However, if swelling, hypotension, or ocular symptoms such as double vision or squint are observed, the identity of the snake is most likely mamushi. In severe cases, nausea, vomiting, stomachache, diarrhea, cyanosis, and tachycardia are sometimes observed.

Grade classification for mamushi bites is clinically used to determine the severity of injuries as follows [17,29]: Grade I, redness and swelling around the bitten area; Grade II, redness and swelling of the wrist or foot joint; Grade III, redness and swelling of the elbow or

| Mamushi | Habu | Yamakagashi |
|---------------------|----------------------------------|---|
| Mamushi grade ≧ III | N/A | Fibrinogen <100 mg/dL |
| | | |
| 2.4%-9.0% | 11% | 0% |
| N/A | 24.2% | N/A |
| | Mamushi grade ≧ III 2.4%–9.0% | Mamushi grade ≥ III N/A 2.4%-9.0% 11% |

Table 3 Indication and incidence of side effects in antivenom

N/A not applicable.

knee joint; Grade IV, redness and swelling of the whole extremity; and Grade V, redness and swelling in parts beyond the extremity or exhibiting systemic symptoms.

Habu (P. flavoviridis)

There are no standardized diagnostic or severity criteria for habu bites. Local swelling may help determine whether the patient was bitten by a habu. Because habu bites result in swelling within 30 min, the circumference of the affected limb may be one indicator of severity. Twenty percent of habu bites are dry. This incidence is higher than that of bites by other snakes, such as the saw-scaled viper (*Echis carinatus*) with 8% dry bites and the rattlesnake in Central California with 10.9% dry bites [30-32]. While most dry bite cases do not require admission, Levine recommends repeating laboratory test within 6 h [4].

Yamakagashi (R. tigrinus)

Yamakagashi bites have been diagnosed based on detailed descriptions of snakes by patients and hemorrhagic symptoms including severe hypofibrinogenemia (<100 mg/dL) [6]. In one study, about 80% of the reported patients developed persistent bleeding from the bite site on admission [5]. DIC with a fibrinolytic phenotype develops early; therefore, evaluating the DIC score is mandatory for the diagnosis of this injury [33]. Assessment of the levels of antithrombin III (AT-III), thrombinantithrombin III complex (TAT) and plasmin- α 2-plasmin inhibitor complex (PIC) may help to evaluate the clinical condition.

Treatment

Previously recommended first-aid measures are strongly discouraged [3]. The use of tight ligatures and arterial tourniquets in the first-aid treatment of snakebite has been universally condemned by modern snakebite experts due to the increase of potential adverse effects and the lack of effectiveness [34-36]. No human study has shown the efficacy of incision and suction as a first-aid tool with regard to improvement of survival or outcome [37].

Once airway, breathing, and circulation have been established, a rapid, detailed history should be obtained [3,37]. If a snake bite is suspected, hospital admission should be considered for further follow-up.

Antivenom therapy

Snake antivenoms are manufactured by immunizing horses against unbound venom. Antivenom treatment is the definitive therapy, but not all cases warrant such therapy (Table 3). Antivenom is administered intravenously to achieve rapid onset of action [7,11]. Subcutaneous or intramuscular injection for the purpose of avoiding side effects is not recommended.

Because snakes inject the same amount of venom into adults and children, the same dose/volume of antivenom must be administered to children.

Preparedness for anaphylaxis should be considered when administering the antivenom. Premedication with an antihistamine and/or epinephrine should be used when the perceived benefit is greater than the risk of adverse effects [5]. As for the use of hydrocortisone as premedication for snake antivenom, the efficacy has not been determined [38].

Another major adverse effect of antivenom is serum sickness disease, which usually occurs 4–10 days after antivenom administration [39]. Rashes, itching, joint pain, fever, lymphadenopathy, malaise, and renal failure are typical symptoms [39,40]. Serum sickness disease is the prototypical type III hypersensitivity reaction, involving excessive immune complex formation [41]. Although many patients have mild symptoms, the reaction can lead to multiple organ failure. Such severe reactions most often occur in patients with severe snakebites that require large amounts of antivenom. Systematic corticosteroids are the main treatment of choice, starting at a dose of 60 mg per day and tapering over 2 weeks to avoid rebound [3,42]. Plasmapheresis is used to obtain rapid effectiveness, particularly in severe cases [43,44].

Mamushi and habu antivenom are approved drugs, whereas yamakagashi antivenom is used as an off-label drug in Japan. Therefore, clinicians are required to join a clinical research group to use yamakagashi antivenom in clinical practice [6].

Efficacy of antivenom

Mamushi (G. blomhoffii)

Studies have evaluated the efficacy of antivenom and cepharanthine (CEP) in a single-center cohort study [45,46]. Makino et al. evaluated 114 cases and reported that patients administered antivenom had a significantly

shorter hospital stay than those administered CEP (p < 0.01). However, in severe cases (grades of mamushi bites IV/V), the percentage of patients administered antivenom was higher than that of patients administered CEP (50% vs. 33%, p = 0.06) [45]. In contrast, Kochi et al. evaluated 50 cases and reported that patients administered antivenom had a significantly longer hospital stay than those administered CEP because of the greater severity of cases in the antivenom group [46]. Thus, evaluating the efficacy of antivenom and CEP without adjusting for the severity of mamushi bites limited these studies [45,46].

Until 1990, antivenom was most often administered subcutaneously or intramuscularly to avoid adverse reactions. Due to its slow absorption in the human body, mamushi antivenom was mistakenly assumed to be ineffective by clinical doctors [47]. Intravenous administration of antivenom had started in the 1990s, and proper reevaluation of the antivenom was expected. Hifumi et al. conducted large, multi-center, population-based studies [17], reporting 234 mamushi bites. Among the severe cases (grades III/IV/V), patients administered antivenom had a significantly shorter hospital stay than those administered CEP (p = 0.024). In contrast, for the mild cases (grades I/II), there was no significant difference in the duration of hospital stay between the two groups (p = 0.77). Therefore, the authors concluded that antivenom is effective in shortening the duration of hospital stay for patients with severe mamushi bites [17]. We propose a new clinical decision algorithm for mamushi bites as shown in Figure 5. We recommend antivenom administration in patients with mamushi grade ≧III on the basis of our previously reported data [17].

Habu (P. flavoviridis)

No definitive indication for the use of antivenom has been provided in clinical practice. Although antivenom is considered effective following habu bites, there are no large-scale studies of the prognosis. Okinawa prefecture is known to have a large population of habu, and the rate of antivenom use is high. There have been no deaths from habu bites in the last 10 years in this area (2004–2013, no deaths in 551 cases) [48]. However, between 1965 and 1969, there were approximately 24 deaths among 1,770 cases in Okinawa due to the lack of antivenom [48]. Therefore, antivenom therapy is currently considered useful for habu bites [49,50].

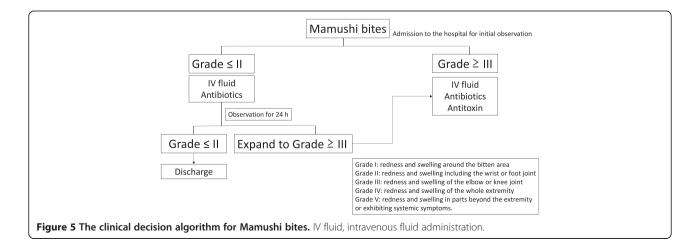
Yamakagashi (R. tigrinus)

Hifumi et al. conducted a retrospective survey analyzing data from 34 patients (19 of whom were treated with antivenom) between 1973 and 2013 [5]. Univariate analysis revealed no significant difference in baseline characteristics and laboratory data between those treated with and without antivenom. Hospital mortality was significantly lower in patients treated with antivenom than in those treated without (0% vs. 26.7%; p = 0.03). Moreover, the number of patients with renal failure requiring hemodialysis was significantly lower among those treated with antivenom (5.3% vs. 40.0%; p = 0.03).

Therefore, antivenom is a specific, definitive, and effective treatment. Administration of yamakagashi antivenom following bites can lead to complete clinical recovery without progression to multiple organ dysfunction syndrome (MODS), even in the presence of severe DIC. Thus, antivenom effectively treats the acute symptoms and can prevent disease progression. Fibrinogen levels <100 mg/dL are considered appropriate for antivenom administration in clinical practice [13,51].

Antivenom side effects Mamushi (G. blomhoffii)

A recent national survey reported that the incidence of adverse reactions to antivenom was 2.4%–9.0%, including mild cases [9,17].



Habu (P. flavoviridis)

Miyagi reported that habu antivenom induced early allergic reactions in approximately 11% and serum sickness disease in approximately 24.2% of patients [7]. The reason this antivenom has higher rates of allergic reactions than the other two antivenoms produced using horses remains unknown.

Yamakagashi (R. tigrinus)

Although the number of the included patients is small (34 cases) to make any comprehensive assessment, the initial anaphylactic reaction rate (including severe reactions) was zero [5,6].

Other treatments

Mamushi (G. blomhoffii)

CEP, a biscoclaurine (bisbenzylisoquinoline) amphipathic alkaloid isolated from Stephania cepharantha Hayata, has been proposed as a possible alternative therapy to antivenom because it lessens the inflammation and pain caused by snake bites [9]. CEP and other extracts from the same plant are widely used in clinical practice (primarily in Japan) to treat a variety of acute and chronic diseases such as alopecia areata [52], radiotherapy-induced leucopenia [53], malaria [54], and septic shock [55]. However, CEP does not have the ability to neutralize circulating venom [56]; therefore, CEP should not be used instead of antivenom for treating mamushi bites (Figure 5). A previously proposed clinical decision algorithm for mamushi bites (supported by the pharmaceutical company) recommends the routine use of CEP [57]; however, the routine use of CEP is clearly unnecessary considering its limited effectiveness.

Because no cases of tetanus associated with mamushi bites have been reported, routine use of tetanus toxoid in patients with mamushi bites is not recommended (Figure 5).

Habu (P. flavoviridis)

Because myonecrosis and CS are often observed, our goal in treatment is not only to save lives but to improve functional outcomes [58,59]. Habu bites caused 14 cases of CS in 2009 [60]. Fasciotomy is required when the compartment pressure reaches 30 mmHg. However, when pressures only moderately exceeds 30 mmHg, some people advocate management with further antivenom, elevation, and reassessment within a few hours; in such cases, fasciotomy is only considered if pressures fail to decrease within several hours [3,32]. This protocol may be the preferable option for mildly symptomatic patients. The initial use of intravenous fluids is also effective for improving circulatory dysfunction and preventing renal dysfunction caused by CS.

Yamakagashi (R. tigrinus)

Yamakagashi bites induce DIC, for which heparin has been used [5]. However, heparin use is contraindicated considering the pathophysiology of DIC involving fibrinolysis. Although other adjunct DIC treatments, such as protease inhibitors and fresh frozen plasma (FFP), are clinically used, the only definitive therapy available is antivenom.

Conclusions

In this review, we have provided information to clarify the clinical diagnosis of snake bites. Antivenom therapy is warranted for venomous snake bites.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

TH, AS, and YK wrote the manuscript. KS, KU, NK, HK, YK, JI, and KK participated and cooperated with the research group. TH, AY, MA, and YK revised and edited the manuscript. All authors read and approved the final manuscript.

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